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| APPLICATION NO.                             | FILING DATE                    | FIRST NAMED INVENTOR | ATTORNEY DOCKET NO.    | CONFIRMATION NO. |
|---|--------------------------------|----------------------|------------------------|------------------|
| 10/525,259                                  | 02/22/2005                     | Margaret Sin Ka Wan  | 13404                  | 5000             |
| 26874<br>FROST BROV                         | 7590 01/13/201<br>VN TODD, LLC | Ī                    | EXAM                   | IINER            |
| 2200 PNC CENTER                             |                                |                      | FERNANDEZ, SUSAN EMILY |                  |
| 201 E. FIFTH STREET<br>CINCINNATI, OH 45202 |                                |                      | ART UNIT               | PAPER NUMBER     |
|   |                                |                      | 1651                   |                  |
|   |                                |                      |                        |                  |
|   |                                |                      | NOTIFICATION DATE      | DELIVERY MODE    |
|   |                                |                      | 01/13/2011             | ELECTRONIC       |

## Please find below and/or attached an Office communication concerning this application or proceeding.

The time period for reply, if any, is set in the attached communication.

Notice of the Office communication was sent electronically on above-indicated "Notification Date" to the following e-mail address(es):

patents@fbtlaw.com

# Office Action Summary

| Application No.    | Applicant(s)         |  |  |
|--------------------|----------------------|--|--|
| 10/525,259         | WAN, MARGARET SIN KA |  |  |
| Examiner           | Art Unit             |  |  |
| SUSAN E. FERNANDEZ | 1651                 |  |  |

|   | SUSAN E. FERNANDEZ   | 1651   |             |
|---|--|--|-------------|
| The MAILING DATE of this communication appears on the cover sheet with the correspondence address Period for Reply  |  |  |             |
| A SHORTENED STATUTORY PERIOD FOR REPLY WHICHEVER IS LONGER, FROM THE MAILING DA Extensions of time may be available under the provisions of 37 CPR 1.13 after SIX (6) MONTHS from the mailing date of this communication.  I IN Operator trengt is generated advove, the maximum statutory period with the provision of | TE OF THIS COMMUNICATION 6(a). In no event, however, may a reply be tin ill apply and will expire SIX (6) MONTHS from cause the application to become ABANDONE   | N.<br>nely filed<br>the mailing date of this o<br>D (35 U.S.C. § 133). |             |
| Status  |  |  |             |
| Responsive to communication(s) filed on 18 Au  2a) This action is FINAL. 2b) This  3) Since this application is in condition for allowan closed in accordance with the practice under E.  | action is non-final.<br>ce except for formal matters, pro  |  | e merits is |
| Disposition of Claims   |  |  |             |
| A  Claim(s) 1.5-7.9-16.18-20.25-30.35.36 and 49-48) Of the above claim(s) is/are withdraw   | n from consideration.  | on.  |             |
| Application Papers  |  |  |             |
| 9) The specification is objected to by the Examiner 10) The drawing(s) filed on is/are: a) acce Applicant may not request that any objection to the c Replacement drawing sheet(s) including the correction 11) The oath or declaration is objected to by the Example.  | epted or b)  objected to by the l<br>drawing(s) be held in abeyance. See<br>on is required if the drawing(s) is obj  | e 37 CFR 1.85(a).<br>jected to. See 37 C                               |             |
| Priority under 35 U.S.C. § 119  |  |  |             |
| 12) 🖾 Acknowledgment is made of a claim for foreign   a) 🖾 All b) □ Some * c) □ None of:  1. □ Certified copies of the priority documents 2. □ Certified copies of the priority documents 3. ☒ Copies of the certified copies of the priori application from the International Bureau * See the attached detailed Office action for a list of   | s have been received. In have been received in Application of the second | on No<br>ed in this National   | Stage       |
| Attachment(s)   |  |  |             |
| Notice of References Cited (PTO-892)  | 4) Interview Summary   | (PTO-413)  |             |

| 1) 🔲 1 | Notice of | References | Cited | (PTO-892) |  |
|--------|-----------|------------|-------|-----------|--|
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Notice of Draftsperson's Patent Drawing Review (PTO-94s)
 Information Disclosure Statement(s) (PTO/SB/08)

Paper No(s)/Mail Date \_

| 4) 🔲 | Intervier | w Summa | ary (PTO-413 |
|------|-----------|---------|--------------|

Paper No(s)/Mail Date. \_\_\_\_.

5) Notice of Informal Patent Application 6) Other: \_\_\_\_\_

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#### DETAILED ACTION

The amendment filed August 18, 2010, has been received and entered.

Claims 2-4, 8, 17, 21-24, 31-34, and 37-48 are cancelled.

Claims 1, 5-7, 9-16, 18-20, 25-30, 35, 36, and 49-56 are pending and examined on the merits to the extent they read on the elected subject matter.

### Claim Rejections - 35 USC § 103

The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:

(a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negatived by the manner in which the invention was made.

Claims 1, 5-7, 9-16, 18-20, 25, 27-30, 35, 36, 50, 51 and 54-56 are rejected under 35 U.S.C. 103(a) as being unpatentable over Sanders et al. (US 2003/0211130).

Sanders et al. teaches the creation of a target tissue substitute wherein a scaffold comprising one or more layers of one or more arrays of microfibers is provided to mimic the configuration of one or more structural elements in a target tissue, and then cells are cultured on the scaffold (page 1, paragraph [0006]). The microfibers can be made of poly-L-lactic acid/polycaprolactone co-polymers, polyglycolic acid (page 1, paragraph [0007]), or poly(lactic acid) (also known as polylactide) (page 5, paragraph [0052]). To form the arrays of microfibers, electrospinning may be used wherein "...a polymer solution held by its surface tension at the end opening of a capillary tube is subjected to an electric field, charge is induced at the liquid

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surface." And then, a fine jet of the polymer solution travels to form fine fibers on a plate, screen, or mandrel. See page 5, paragraph [0050]. The polymer solution for electrospinning may also be a polymer melt (page 5, paragraph [0051]). Thus, Sanders et al. teaches supplying a liquid solution containing a biologically compatible polymer dissolved in a liquid to a liquid outlet placed in the vicinity of a surface, subjecting the liquid solution from the outlet to an electric field to form polymer fibres on a surface.

As an array of microfibers is formed that may be arranged in multiple layers (page 1, paragraph [0009]), where the microfibers can be oriented at a defined angle with respect to adjacent fibers (page 5, paragraph [0057]), and the microfibers are designed to mimic the configuration of one or more structural elements in a target tissue (page 1, paragraph [0006]), Sanders et al. indeed teaches the creation of a three-dimensional continuous network of intercommunicating fibre portions with gaps between adjacent fibre portions. For instance, page 14, paragraph [0141] teaches the formation of a "dense elastin network." Clearly the resulting fibre scaffold is a "network-like formation" as required by the instant claims. Moreover, given that cells are applied to the fibre scaffold to form a target tissue substitute (page 1, paragraph [0006]), the cell processes such as attachment, movement, growth, proliferation, and differentiation are facilitated by the fibre scaffold. Note that the cells cultured on the scaffold may be fibroblast cells (page 12, paragraph [0116]).

Sanders et al. also teaches that the microfibers have a diameter between about 1 micrometer to about 20 micrometers (page 1, paragraph [0007]) and that the gaps between adjacent microfibers range from about 10 micrometers to 100 micrometers (page 1, paragraph [0008]), thus meeting the size limitations recited in instant claims 1, 14, 20, and 54.

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Sanders et al. differs from the claimed invention in that it does not specify that the cells are mammalian cells such as human bone marrow fibroblastic cells, human adherent cells, and stem cells, the cell diameters, the relationship between the mammalian cell diameter and the fibre diameter or the fibre gap, or the relationship between the size of the cell surface receptors of the cells and the fibre diameter. However, it would have been obvious to have used any type of cell, including human bone marrow fibroblastic cells, human adherent cells, and stem cells, since the scaffold created by Sanders et al. may be provided for the different tissues and organs in the body (page 12, paragraph [0117]). In using a variety of cells, a wide range of cell diameters are obvious, and thus the cell diameters can be from about 5 to about 10 times greater than the fibre diameter and/or the fibre gap is greater than approximately half the cell diameter. Moreover, in rendering obvious the relationship between the cell diameters, the fibre diameters, and the fibre gaps, cell processes including attachment, movement, growth, proliferation, and differentiation are inherently facilitated. Thus, instant claims 1, 5-7, 9, 11-15, 20, 25, 27, 35, 36, 50, and 54 are rendered obvious.

Furthermore, it would have been obvious to have used a variety of fibre diameters and fibre gap sizes, including those recited in instant claims 16 and 18, since it would have been a matter of routine experimentation and optimization. Thus, instant claims 16, 18, 19, and 50 are rendered obvious.

Also, it would have been obvious to have used different types of polylactide, including those recited in instant claim 10, since Sanders et al. broadly teaches the used of polylactide. There would have been a reasonable expectation of success in substituting one polylactide for another. Thus, instant claim 10 is rendered obvious.

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Additionally, it would have been obvious to have used a variety of solvents, including cell culture medium, water, acetone, ethanol, and DMEM, since they are known solvents, are deemed suitable for cell growth, and would allow for the formation of a polymer solution. Thus, instant claims 29, 55, and 56 are rendered obvious.

With respect to instant claims 28 and 51, it would have been obvious to have used any known method, including spraying, to apply the cells to the scaffold. There would have been a reasonable expectation of success is applying the cells by spraying since the cells can be present in a solution. Thus, instant claims 28 and 51 are rendered obvious. Also, it would have been obvious to have added the cells to the polymer solution prior to supplying the polymer solution to the outlet since there would have been a reasonable expectation of success in applying the cells at any point of the preparation of the scaffold with cells. Thus, instant claim 30 is rendered obvious.

A holding of obviousness is clearly required.

Claims 1, 5-7, 9-16, 18-20, 25-30, 35, 36, and 49-56 are rejected under 35 U.S.C. 103(a) as being unpatentable over Sanders as applied to claims 1, 5-7, 9-16, 18-20, 25, 27-30, 35, 36, 50, 51 and 54-56 above, and further in view of Coffee et al. (WO 98/03267).

As discussed above, Sanders render claims 1, 5-7, 9-16, 18-20, 25, 27-30, 35, 36, 50, 51 and 54-56 obvious. However, they do not expressly disclose that the fibers are directly applied to a target area of a mammalian body, such as a wound, to form the fibre scaffold in situ.

Coffee et al. discloses a method of depositing fibres on a surface, such as a wound on an animal to form a dressing (page 4, second paragraph). A solution comprising a biocompatible polymer is subjected to an electrohydrodynamic process in the vicinity of the surface (page 4, third paragraph).

At the time the invention was made, it would have been obvious to the person of ordinary skill in the art to have applied the fibers directly to a wound on a mammalian body to form the fibre scaffold of the Sanders invention in situ. One of ordinary skill in the art would have been motivated to do this since it would have resulted in a dressing for the wound and since the application of a electric field is suitable for depositing fibres on a wound. Thus, instant claims 26, 49, 52, and 53 are rendered obvious.

#### Response to Arguments

Applicant's arguments filed August 18, 2010, have been fully considered but they are not persuasive. The applicant argues that Sanders only teaches the use of linear arrays, and that the use of nonlinear, mesh-type arrays is discouraged. Paragraph 138 of Sanders, which describes the results of Example 1, is pointed to as support for the applicant's assertion that Sanders teaches away from the claimed "fibre scaffold comprising a lattice or network-like formation." Example 1 compares arrays of microfibers with an electrospun fibro-porous mesh as a control (page 13, paragraph [0130]). Sanders points out that "Excessive spacing may results in gaps or holes where cells do not grow into confluency. Thus, the overall architecture of the scaffold may affect the organization of the tissue structure. For example, the use a mesh-like scaffold may result in a disorganized tissue structure with respect to the orientation of cells, as well as the protein synthesized by the cells (for example, collagen), as shown in Example 1," (page 6, paragraph [0058]). Clearly, it is not that the shape (mesh) of the mesh-type array of Example 1

that disadvantageously results in the disorganized cell orientation, but the excessive spacing between the fibres of the mesh-type array. Therefore, paragraph [00138] of Sanders cannot be regarded as a teaching away of the claimed fibre scaffold comprising a lattice or network-like formation. Moreover, Example 2 even teaches the Sanders scaffold as a network of fibres, specifically indicating a "dense elastin network" (page 14, paragraph [0141]). Therefore, the claimed invention must remain rejected over Sanders.

No claims are allowed.

Applicant's amendment necessitated the new ground(s) of rejection presented in this

Office action. Accordingly, THIS ACTION IS MADE FINAL. See MPEP § 706.07(a).

Applicant is reminded of the extension of time policy as set forth in 37 CFR 1.136(a).

A shortened statutory period for reply to this final action is set to expire THREE MONTHS from the mailing date of this action. In the event a first reply is filed within TWO MONTHS of the mailing date of this final action and the advisory action is not mailed until after the end of the THREE-MONTH shortened statutory period, then the shortened statutory period will expire on the date the advisory action is mailed, and any extension fee pursuant to 37 CFR 1.136(a) will be calculated from the mailing date of the advisory action. In no event, however, will the statutory period for reply expire later than SIX MONTHS from the date of this final action.

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Any inquiry concerning this communication or earlier communications from the examiner should be directed to SUSAN E. FERNANDEZ whose telephone number is (571)272-3444. The examiner can normally be reached on Mon-Fri 8:30 am - 5:00 pm.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Mike Wityshyn can be reached on (571) 272-0926. The fax phone number for the organization where this application or proceeding is assigned is 571-273-8300.

Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see http://pair-direct.uspto.gov. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free). If you would like assistance from a USPTO Customer Service Representative or access to the automated information system, call 800-786-9199 (IN USA OR CANADA) or 571-272-1000.

/Allison M. Ford/ Primary Examiner, Art Unit 1651 Susan E. Fernandez Examiner Art Unit 1651

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